

(REVIEW ARTICLE)



## Family Nycteribidae (Insecta: Diptera)

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### Abstract

Family Nycteribidae bat flies have an interesting reproductive method known as viviparous puparity, in which eggs are fertilized internally and all three larval stages develop within the female. Larvae are nourished by intrauterine glands. Gravid female flies deposit a single, 3rd instar larva on the roosting substrate. The larva then immediately forms a puparium, and following approx. After 3–4 weeks of development, the adult fly emerges to locate and colonize a host. The aim of this article was to investigate the morphological, bioecological, and systematic characteristics of the family Nycteribidae (Insecta: Diptera). For the review, we used indexed articles, scientific book chapters, theses databases, university dissertations, national and international scientific articles, scientific journals, documents, and academic and scientific journals available online ResearchGate, HAL SSRN, Scielo, and Qeios. The present work uses the reference of bibliographical research, understood as the act of inquiring and seeking information on a certain subject, through a survey carried out in national and foreign databases, with the objective of detecting what exists of consensus or controversy.

**Keywords:** Bats; Dipteran; Ectoparasitic; Larvae; Viviparous

### 1. Introduction

The superfamily Hippoboscoidea has listed four families of Calyptrate flies, Glossinidae, Hippoboscidae, Nycteribiidae, and Streblidae. Glossinidae comprises hematophagous flies restricted to the Ethiopian Region, while the other three families include hematophagous flies, ectoparasites of birds, and mammals with a cosmopolitan distribution. Hippoboscidae are fly ectoparasites of birds and mammals (artiodactyls, lemurs, and kangaroos). Nycteribiidae and Streblidae are formed by exclusively ectoparasitic species of bats [1-2].

#### *Objective*

The aim of this article was to investigate the morphological, bioecological, and systematic characteristics of the family Nycteribidae (Insecta: Diptera).

### 2. Methods

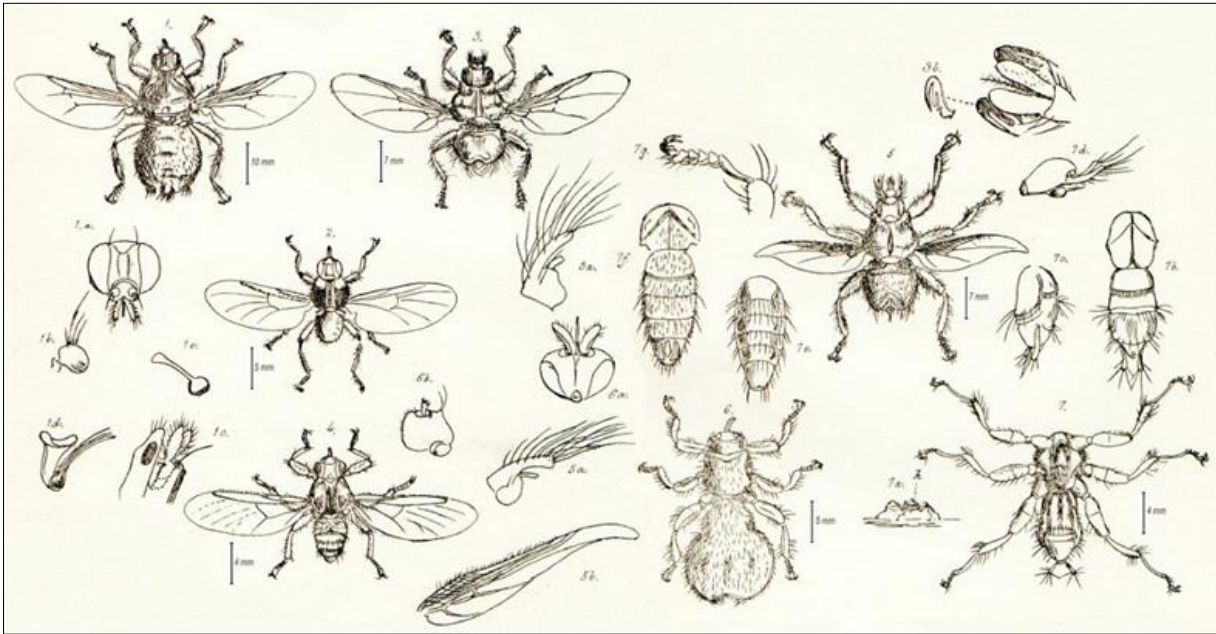
For the review, we used indexed articles, scientific book chapters, theses databases, university dissertations, national and international scientific articles, scientific journals, documents, and academic and scientific journals available online ResearchGate, HAL SSRN, Scielo, and Qeios. The present work uses the reference of bibliographical research, understood as the act of inquiring and seeking information on a certain subject, through a survey carried out in national and foreign databases, with the objective of detecting what exists of consensus or controversy.

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### 3. Family Nycteribidae

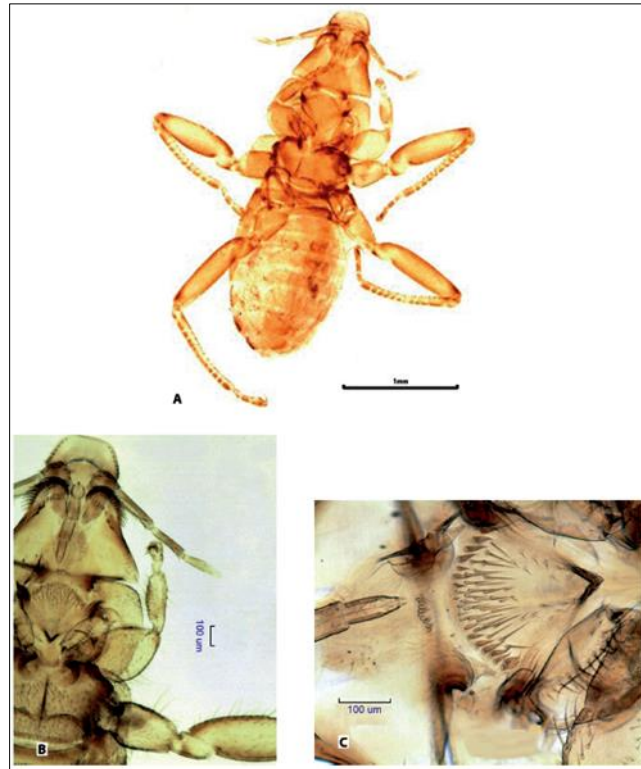
The bat is a mammalian animal of the order Chiroptera distinguished from birds, as these have feathers supported by bones. Bats are the only mammals with true flight. Bats have the most varied diet among mammals, as they can eat fruits, seeds, leaves, nectar, pollen, arthropods, small vertebrates, fish, and blood [1-3].

About 70% of bats are insectivores, feeding on insects, and practically all the rest are frugivorous, that is, they feed on fruits. Only three species feed exclusively on blood: they are the so-called hematophagous or vampire bats, found only in Latin America. In this way, bats contribute substantially to the structure and dynamics of ecosystems, since they act as pollinators, seed dispersers, insect predators (including agricultural pests), nutrient suppliers in caves, and vectors of wild diseases, among other functions. They also have an extraordinary sense of echolocation which they use for orientation, search for food and communication (Figures 1-2) [1-4].



Source: <https://www.delta-intkey.com/britin/dip/www/nycterib.htm>

**Figure 1** *Hippobosca equina* Linnaeus, 1758, with details of head from the front (1a), antenna (1b), front of head from the side (1c), labrum, with lingua and labium and the muscular base by which they are protruded (1d), and one of the halteres (1e). 2 and 4, *Hippobosca cervi* Olivier, 1792. 3, *Ornithomya avicularia*. (Linnaeus, 1758), with details of antenna (3a), and tarsal claws (3b). 5, *Ornithomya pallida* Latreille, 1812. 5a and 5b, *Stenopteryx hirundinis* (Linnaeus, 1758), antenna and wing respectively. 6, *Melophagus ovinus* (Linnaeus, 1758), with details of head from beneath (6a), and antenna (6b). Nycteribidae. 7, *Phthiridium biarticulatum* Hermann, 1804; 7a, the same, with head (h) thrown back in the normal position; 7b, the thorax and abdomen from beneath; 7c and 7d, the same, showing abdomen from the side (7c) and the porrected head from the side (7d). 7e and 7f, *Nycteribia kolenatii* Theodor, 1954: abdomen from above (7e), and thorax and abdomen from below (7e). From Walker (1853), with approximate lengths of insects (front of head to tip of abdomen)

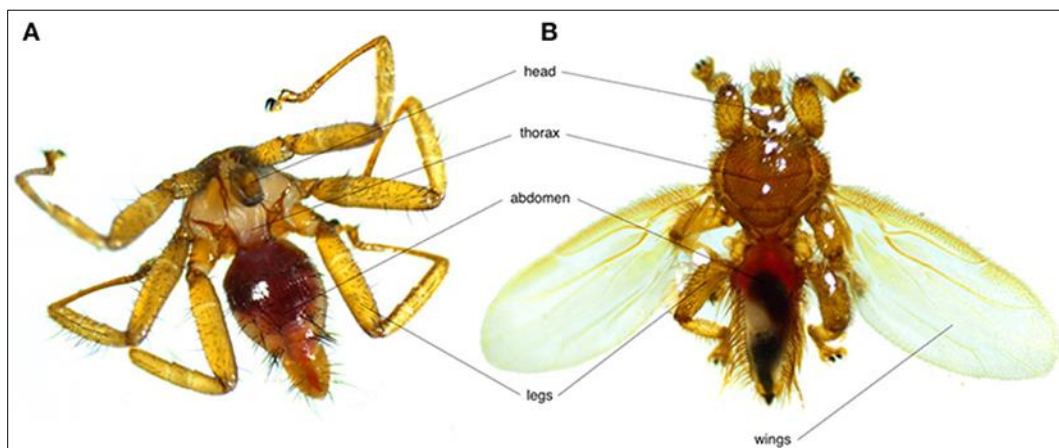


Source: <https://www.scielo.br/j/paz/a/spcpTn66ZsVXbmWRdYz7pJC/?lang=en#ModalFig4>

**Figure 2** *Hesperoctenes cartus* Jordan, 1922 (CMLA 766 ♀): (A) Ventral view, (B) Antenna and anterior leg, (C) Gular region and Prosternum

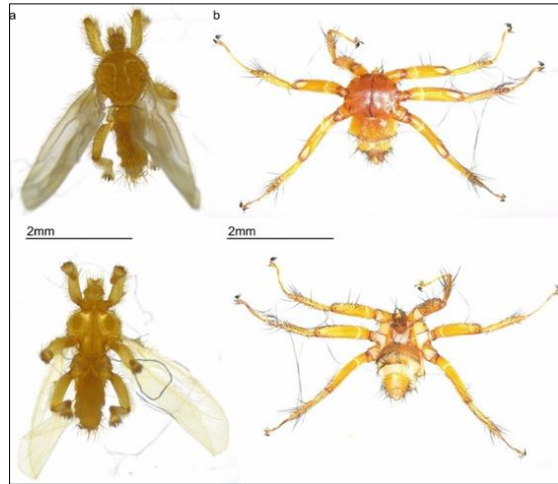
### 1.1. Description

The species have the mesonotum reduced and depigmented and the pleurae displaced to the dorsal surface due to expansion of the sternum thoracic, making them look acariform. O body length ranges from 1.5 to 5 mm. As in the other families of Hippoboscoidea, the development of the larva takes place in the mother's uterus, where it is fed through glands that produce a nutritive substance. In the third instar, the larva is expelled and deposited in the shelter substrate where it passes to the pupal stage (Figures 3-7) [5-7].



Source: <https://www.frontiersin.org/articles/10.3389/fvets.2019.00115/full>

**Figure 3** Photos showing the morphological differences between (A) a wingless nycteribiid and (B) a streblid bat fly



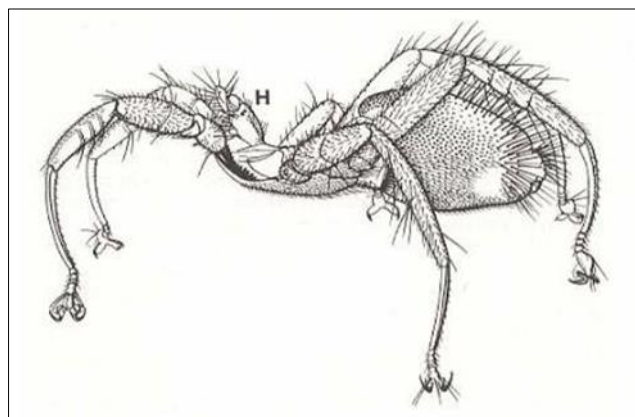
Source: [https://www.researchgate.net/figure/Typical-morphology-of-bat-flies-belonging-to-the-family-a-Streblidae-and-b-Nycteribiidae\\_fig2\\_354875976](https://www.researchgate.net/figure/Typical-morphology-of-bat-flies-belonging-to-the-family-a-Streblidae-and-b-Nycteribiidae_fig2_354875976)

**Figure 4** The head is the hairy ovoid structure inserted after the 2 front legs and is conspicuously small with either no eyes or very small ones. And just look at their legs – amazingly well adapted for holding onto the fur of bats. Their tarsal segments are completely bendy with huge claws



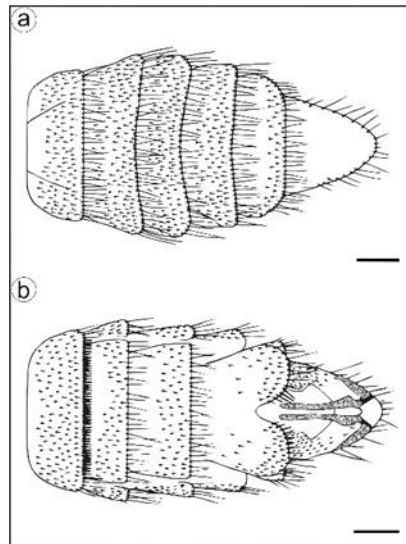
Source: [https://www.nhm.ac.uk/natureplus/blogs/diptera-blog/tags/batlice\\_flies.html](https://www.nhm.ac.uk/natureplus/blogs/diptera-blog/tags/batlice_flies.html)

**Figure 5** Their legs are amazingly well adapted for holding onto the fur of bats



Source: [https://www.nhm.ac.uk/natureplus/blogs/diptera-blog/tags/batlice\\_flies.html](https://www.nhm.ac.uk/natureplus/blogs/diptera-blog/tags/batlice_flies.html)

**Figure 6** The batlice fly has an extremely small thorax. So the head and legs are basically dorsal insertions of the thorax – they stick out of the top of the fly rather than at the side which is the more usual way. It is really difficult to work out which way is up as the head does not resemble anything that you are used to

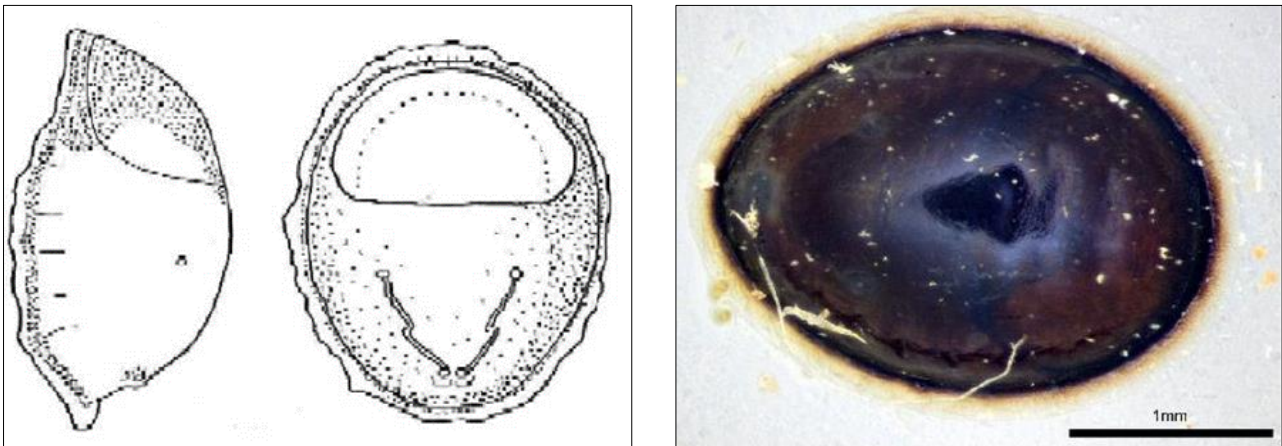


Source: <https://www.scielo.br/j/aabc/a/57vTmRfRgWB8f6xjCcXZfZP/>

**Figure 7** a) Abdomen, female (Holotype), dorsal view, *Hershkovitzia mariae* sp. nov. b) Abdomen, female (Holotype), ventral view, *H. mariae*

### 1.2. Biology

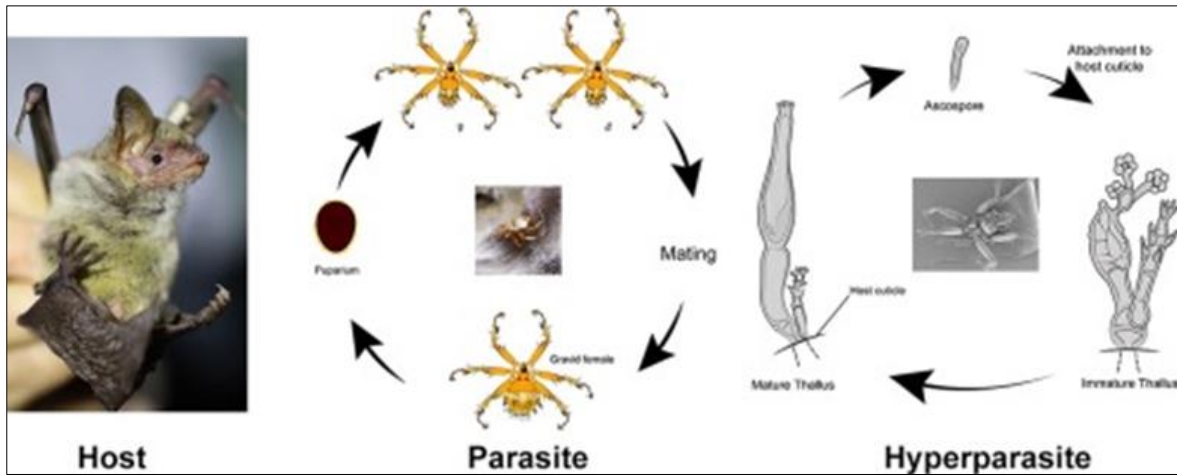
Larvae and pupae. The larvae develop within the mother, being nourished via a greatly developed ‘accessory gland’, before leaving to pupate (finally pupating on the ground or in the abode of the host); acephalic. The pupae are enclosed within a puparium (Figure 8).



Source: [https://www.nhm.ac.uk/natureplus/blogs/diptera-blog/tags/batlice\\_flies.html](https://www.nhm.ac.uk/natureplus/blogs/diptera-blog/tags/batlice_flies.html)

**Figure 8** The females leave the bat host when they are about to give birth (as it were). She crawls onto the wall of the cave and the pre-pupal stage emerges. This is an incredibly short stage as the larva pupates within hours. Now there are some great larval body adaptations to help this wee one stick to the cave wall. They are hemi ovoid and have sticky secretions which are also helped by a narrow marginal skirt

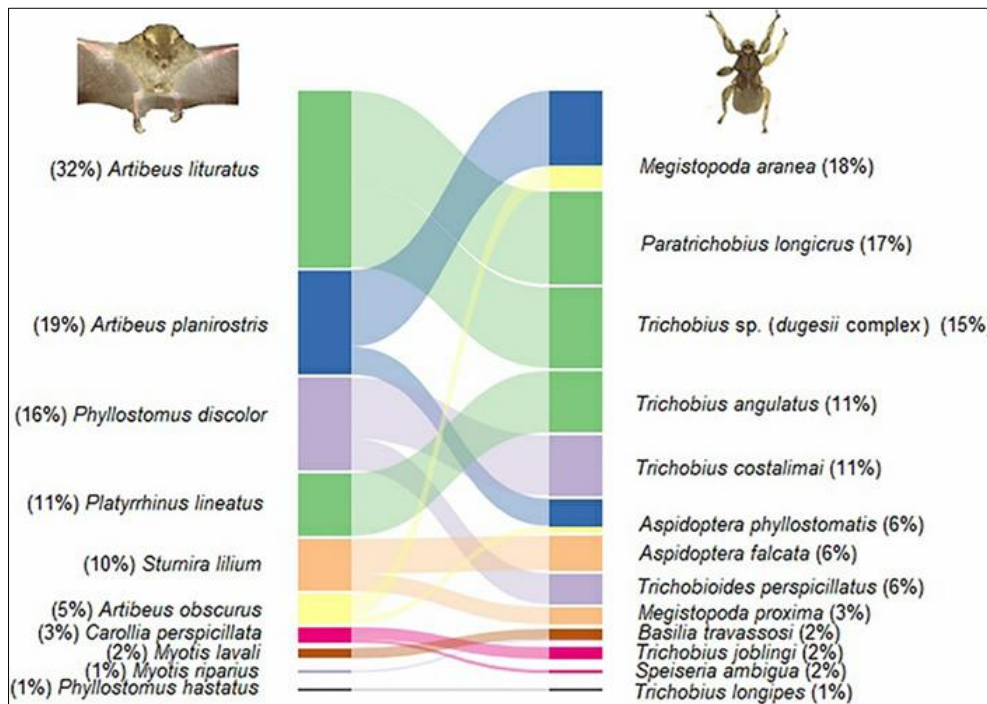
Given the dorsal insertion of the legs and general morphology, nycteribiid flies generally move well in any direction, and their movements may be very fast when agitated. This high mobility in the bat's fur may afford them some protection from host grooming, which is understood to be a primary cause of adult fly mortality (Figure 9) [8-10].



Sources: <https://doi.org/10.1016/j.jip.2019.107206> <https://www.sciencedirect.com/science/article/abs/pii/S0022201119301193>

**Figure 9** Bat flies (Diptera: Nycteribiinae) are highly specialized bloodsucking bat ectoparasites. Some of the ectoparasitic bat flies are themselves parasitized with an ectoparasitic fungus of the genus *Arthrorhynchus* (Laboulbeniales). Ascospores of the fungus attach to the cuticle of a bat fly and develop a haustorium that penetrates the host cuticle. This interaction defines the fungus as a hyperparasite. Both the fly and the fungus are obligate parasites and this peculiar case of hyperparasitism. *Laboulbeniales*, Engler (1898), genus *Arthrorhynchus* Kolen, in natural populations of bat flies infesting the bat species *Miniopterus schreibersii* (Natterer, 1819), *Myotis bechsteinii*, *Myotis bechsteini* (Kuhl, 1817), *Myotis daubentonii* (Kuhl, 1817), *Myotis escalerae* Cabrera, 1904 and *Myotis* sp. in Portuguese cave

Bat flies have an interesting reproductive method known as viviparous puparity, in which eggs are fertilized internally and all three larval stages develop within the female. Larvae are nourished by intrauterine glands. Gravid female flies deposit a single, 3rd instar larva on the roosting substrate. The larva then immediately forms a puparium, and following approx. 3–4 wk development, the adult fly emerges to locate and colonize a host (Figure 10) [11-13].



**Figure 10** Interaction network between bats and ectoparasites captured in urban green areas of Grande Aracaju, Sergipe, Brazil. The lines and their different colors represent interactions between species, and the width of the line indicates the strength of the interactions. The values in parentheses indicate the frequency of occurrence of the species

### 1.3. Classification

Currently, the family comprises 260 species, including 13 genera and three subfamilies, Nycteribiinae, Cyclopodiinae, and Archinycteribiinae. On the American continent, two genera are found Nycteribiinae, *Basilina* Miranda-Ribeiro, 1903, *Herskovitzia* Guimaraes & D'Andretta, 1956, are endemic to the Neotropical Region and are four known species that parasitize bats of the family Thyropteridae. In Brazil, there were two species recorded, *Herskovitzia cabala* Peterson & Lacey, 1985 and *Herskovitzia inaequalis* Theodor, 1967, both records in the Amazon region. *Basilina* is the genus with the highest number of species within Nycteribiidae, currently over 100, 47 of which are exclusively American. The species are found on all continents and mainly parasitize bats of the family Vespertilionidae. Some Streblidae and Nycteribiidae (Diptera: Hippoboscoidea) from Maracá Island, Roraima, Brazil [14-17].

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## 4. Selected Manuscripts

### 4.1. Study 1

Some Streblidae and Nycteribiidae (Diptera: Hippoboscoidea) from Maracá Island, Roraima, Brazil.

4.1.1. *Basilina dunnii* Curran, 1935.

**Hosts:** *Myotis nigricans* (Schinz, 1821) and *Myotis albescens* Geoffroy (Guimarães 1966).

**Distribution:** Panamá and Venezuela.

4.1.2. *Basilina ferrisi* Schuurmans-Stekhoven Jr., 1931.

**Hosts:** *Basilina ferrisi* Schuurmans-Stekhoven, 1931, occurs on *Myotis albescens* (Geoffroy, 1806), *M. nigricans*, and *Myotis riparius* Handley, 1960.

**Distribution:** Costa Rica, Colômbia, Guatemala, Guiana, Peru, and Venezuela [18].

### 4.2. Study 2

- Nycteribiidae Samouelle, 1819.
- *Basilina* Miranda-Ribeiro, 1903.
- *Pseudelytromyia* Miranda-Ribeiro, 1907.
- *Basilina andersoni* Peterson & Maa, 1970.

**Hosts.** *M. nigricans* (Vespertilionidae), *Eptesicus brasiliensis* (Desmarest, 1819), *Myotis* sp. and *Histiotus velatus* (Geoffroy, 1824) (Vespertilionidae).

**Distribution:** Brazil (Parana and Rio Grande do Sul) and Uruguay.

4.2.1. *Basilina carteri* Scott, 1936.

**Hosts:** *Molossops temminckii* (Burmeister, 1854) (Molossidae), *Eptesicus brasiliensis* (Desmarest 1819), *Eptesicus diminutus* Osgood, 1915, *Eptesicus furinalis* (d'Orbigny & Gervais, 1847), *Myotis albescens* (Geoffroy, 1806), *Eptesicus keaysi* Allen, 1914, *M. nigricans*, *Myotis riparius* Handley, 1960, *Myotis ruber* (Geoffroy, 1806), *Myotis* sp., (Vespertilionidae); *Molossus molossus* Pallas, 1766 e *Tadarida brasiliensis* (Geoffroy, 1824) (Molossidae).

**Distribution:** Brazil (Mato Grosso, São Paulo, Paraná e Santa Catarina), Bolívia, Paraguay and Argentina (Jujuy, Tucumán, Salta and Santiago del Estero).

4.2.2. *Basilina currani* Guimarães, 1943.

**Hosts:** *Myotis ruber* (Vespertilionidae), *E. brasiliensis*, *M. albescens*, *Myotis chiloensis* (Waterhouse 1840), *M. levis* (Vespertilionidae) and *T. brasiliensis* (Molossidae).

**Distribution:** Brazil (São Paulo and Santa Catarina) and Argentina (Catamarca, Tucumán and La Rioja).

**Hosts:** *Basilina hughscotti* Guimarães, 1946, *Chrotopterus auritus* (Peters, 1856) (Phyllostomidae), *E. furinalis*, *M. nigricans*, and *M. riparius* (Vespertilionidae).

**Distribution:** Brazil (Distrito Federal, Minas Gerais and Rio Grande do Sul).

*Basilina lindolphoi* Graciolli, 2001.

**Distribuição geográfica.** Brazil (São Paulo, Paraná and Santa Catarina).

4.2.3. *Basilina speiseri* (Miranda-Ribeiro, 1907).

**Hosts:** *Lasiurus borealis* (Müller, 1776) (= *Lasiurus blossevillii*) (Vespertilionidae), *Anoura geoffroyi* Gray, 1838, *Carollia perspicillata* (Linnaeus, 1758), *Phyllostoma* sp. (Phyllostomidae); *E. brasiliensis*, *M. albescens*, *M. nigricans* (Vespertilionidae) and *M. obscurus* (Molossidae).

**Distribution:** Brazil (Mato Grosso, Rio de Janeiro, São Paulo, Paraná and Santa Catarina), Paraguay and Argentina (Santa Fé).

4.2.4. *Basilina juquiensis* Guimarães, 1946.

**Hosts:** *M. nigricans*, *M. riparius*, and *Myotis* sp. (Vespertilionidae).

**Distribution:** Venezuela and Brazil (São Paulo, Paraná and Santa Catarina).

4.2.5. *Basilina ferruginea* Miranda-Ribeiro, 1903.

**Hosts:** *Atalapha frantzii* Dobson, 1878 (= *Lasiurus blossevillii*) (Vespertilionidae), *Lasiurus borealis* (Müller, 1776), *Lasiurus cinereus* (Palisot de Beauvois, 1796) and *Lasiurus pfeifferi* (Gundlach, 1861) (Vespertilionidae).

**Distribution:** Cuba, Panamá, Equador (Galápagos); Brazil (Pará, Minas Gerais, Rio de Janeiro, São Paulo and Santa Catarina) and Paraguai.

4.2.6. *Basilina ortizi* Machado-Allison, 1963.

**Hosts:** *Eptesicus melanopterus* (Jentink, 1904) (= *Eptesicus fuscus*) (Vespertilionidae), *Artibeus hartii* Thomas, 1892, *Chrotopterus auritus* (Peters, 1856) (Phyllostomidae); *E. brasiliensis*, *E. furinalis*, *Eptesicus* sp. and *M. riparius*.

**Distribution:** Belize, Costa Rica, Venezuela and Brazil (Paraná).

4.2.7. *Basilina plaumanni* Scott, 1940.

**Hosts:** *Histiotus velatus* (Geoffroy, 1824) (Vespertilionidae), *E. brasiliensis*, *E. fuscus*, *Histiotus macrotus* (Poepfig, 1835), *Histiotus montanus* (Philippi & Landbeck, 1861), *Histiotus* sp. and *M. nigricans*.

**Distribution:** Brasil (Minas Gerais, São Paulo, Paraná, Santa Catarina e Rio Grande do Sul), Paraguai, Argentina (Jujuy, Santiago del Estero and Córdoba) and Uruguai.

**Hosts:** *Myotis albescens* (Vespertilionidae). *M. nigricans* (Vespertilionidae).

**Distribution:** Brasil (Paraná and Santa Catarina).

4.2.8. *Basilina ruiae* Graciolli, 2003.

**Hosts:** *Myotis ruber* (Vespertilionidae).

**Distribution:** Brazil (São Paulo, Paraná and Rio Grande do Sul) [19].

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## 5. Conclusion

Bat flies (Diptera: Nycteribiinae) are highly specialized bloodsucking bat ectoparasites. Some of the ectoparasitic bat flies are themselves parasitized with an ectoparasitic fungus of the genus *Arthrorhynchus* (Laboulbeniales). Ascospores of the fungus attach to the cuticle of a bat fly and develop a haustorium that penetrates the host cuticle. This interaction defines the fungus as a hyperparasite. Both the fly and the fungus are obligate parasites and this peculiar case of hyperparasitism.



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